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FOR: BETA-SHEET BREAKING PEPTIDES

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Sir:

In the matter of the above-identified application for patent, notice is hereby given that the applicant claims as priority:

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Certified copies of the corresponding Convention application(s) were submitted to the International Bureau in PCT Application No. PCT/EP03/50287. Receipt of the certified copy(s) by the International Bureau in a timely manner under PCT Rule 17.1(a) has been acknowledged as evidenced by the attached PCT/IB/304.

Respectfully submitted, OBLON, SPIVAK, McCLELLAND, MAIER & NEUSTADT, P.C.

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Bescheinigung

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Attestation

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Die angehefteten Unterlagen stimmen mit der ursprünglich eingereichten Fassung der auf dem nächsten Blatt bezeichneten europäischen Patentanmeldung überein. The attached documents are exact copies of the European patent application described on the following page, as originally filed.

Les documents fixés à cette attestation sont conformes à la version initialement déposée de la demande de brevet européen spécifiée à la page suivante.

Patentanmeldung Nr. Patent application No. Demande de brevet nº

02100787.7

PRIORITY DOCUMENT

SUBMITTED OR TRANSMITTED IN COMPLIANCE WITH RULE 17.1(a) OR (b)

Der Präsident des Europäischen Patentamts; Im Auftrag

For the President of the European Patent Office

Le Président de l'Office européen des brevets

R C van Dijk



European Patent Office Office européen des brevets



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Si aucun titre n'est indiqué se referer à la description.)

BETA-SHEET BREAKING PEPTIDES

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B-Sheet breaking peptides

Field of Invention

The invention relates to the field of β -sheet breaking peptides, particularly their use in the treatment of diseases such as Alzheimer's disease, Down's syndrome, Dementia pugilistica (including head trauma), Hereditary Cerebral Haemorrhage with amyloidosis of the Dutch type (HCHWA-D) and vascular dementia with amyloid angiopathy.

Background of the Invention

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Alzheimer's disease (AD), first described by the Bavarian psychiatrist Alois Alzheimer in 1907, is a progressive neurological disorder that begins with short-term memory loss and proceeds to disorientation, impairment of judgment and reasoning and, ultimately, dementia. The course of the disease usually leads to death in a severely debilitated, immobile state between four and 12 years after onset. AD has been estimated to afflict 5 to 11 percent of the population over age 65 and as much as 47 percent of the population over age 85. The societal cost for managing AD is very high, primarily due to the extensive custodial care required for AD patients. Currently, there is no treatment that significantly retards the progression of the disease. Several groups have reviewed AD^{1,2}.

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Pathologically, AD is characterized by the presence of distinctive lesions in the victim's brain, revealed on autopsy. These brain lesions include abnormal intracellular filaments called neurofibrillary tangles (NTFs) and extra cellular deposits of amyloidogenic proteins in senile, or amyloid, plaques. Amyloid deposits are also present in the walls of cerebral blood vessels of AD patients. The major protein constituent of amyloid plaques has been identified as a 4.3 kiloDalton peptide called β -amyloid peptide (A β)^{3, 4}. Diffuse deposits of

A β are frequently observed in normal adult brains, whereas AD brain tissue is characterized by more compacted, dense-core β -amyloid plaques.⁵ These observations suggest that A β deposition precedes, and contributes to, the destruction of neurons that occurs in AD^{6,7,8}. In further support of a direct pathogenic role for A β , β -amyloid has been shown to be toxic to mature neurons, both in culture and *in vivo* ^{9,10,11,12}.

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Patients with hereditary cerebral haemorrhage with amyloidosis-Dutch-type (HCHWA-D), which is characterized by diffuse β -amyloid deposits within the cerebral cortex and cerebrovasculature, have been shown to have a point mutation that leads to an amino acid substitution within $A\beta$.¹³

Aβ has also been implicated in vascular dementia with amyloid angiopathy^{14,15,16} and dementia pugilistica.^{17,18}

- Natural Aβ is derived by proteolysis from a much larger protein called the amyloid precursor protein (APP)^{19,20,21,22,23}. The APP gene maps to chromosome 21, thereby providing an explanation for the β-amyloid deposition seen at an early age in individuals with Down's syndrome, which is caused by trisomy of chromosome 21^{24, 25,26}.
- Naturally-occurring Aβ, derived from proteolysis of APP, is 39 to 43 amino acid residues in length, depending on the carboxyl-terminal end point, which exhibits heterogeneity. The predominant circulating form of Aβ in the blood and cerebrospinal fluid of both AD patients and normal adults is Aβ1-40^{27,28}. However, Aβ1-42 and aβ1-43 are also found in β-amyloid plaques ^{29,30,31}

Considerable evidence has accumulated that the pathogenicity of A β results from a change in protein conformation^{32,33}. It is believed that a critical event leading to pathology in Alzheimer's disease, Vascular dementia with amyloid angiopathy and HCHWA-D is the

refolding of a natural and non-pathogenic protein, to yield a pathogenic form. The refolding alters the secondary and tertiary structure of the protein without changing its primary structure.

Amyloid is a generic term that is applied to fibrillar aggregates that have a common structural motif: a β -pleated sheet conformation.^{34,35} These aggregates exhibit special tinctorial properties, including the ability to emit a green birefringent glow after staining with Congo red, and the capacity to bind the fluorochrome thioflavin S^{36,37}. These tinctorial properties form the basis of assays used to detect β -amyloid deposits.

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One approach to the treatment and prevention of Alzheimer's disease has been to develop short peptides having some sequence homology to the natural protein sequence believed to be involved in amyloid formation, but also having one or more amino acids that disfavour or destabilise the formation of β -pleated sheet conformations. The peptides prevent the aggregation of β -amyloid, and thereby prevent its cytotoxic effects. This approach has been suggested in Alzheimer's disease and in prion-related disorders^{38,39,40,41,42}, and has lead to the β -sheet breaking peptides shown below, amongst others:

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US 6,319,498 (Praecis Pharmaceuticals) proposes β -sheet breaking peptides based on A β , and exemplifies amino-terminally-biotinylated peptides. US 6,303,567 (Praecis Pharmaceuticals) proposes peptides based on the β -amyloid peptide, but consisting entirely of D-amino acids, as β -sheet breaking peptides.

While the known β -sheet breaking peptides have provided valuable information and may have utility in treating Alzheimer's disease, the development of peptide drugs is severely limited by the fact that natural peptides are subject to degradation and rapid metabolism in the gut, the liver and in circulation. Furthermore, the desired site of action for treatment of many amyloid-related disorders is in the brain, and peptides, like many other molecules, may have difficulty penetrating the blood brain barrier. The brain itself is also replete with peptidases, which degrade peptide molecules.

Summary of the invention

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It is an object of the invention to provide a β -sheet breaking peptide with improved pharmacological profile.

In a first aspect, the invention provides a compound of the general formula I:

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wherein:

R¹ is selected from C₂₋₆-acyl (optionally substituted by amino, C₁₋₆alkyl amino or di-C₁₋₆alkylamino), and C₁₋₆alkyl (optionally substituted by amino, C₁₋₆alkyl amino or di-C₁₋₆alkylamino);

 R^6 is $NR^{12}R^{13}$ (wherein R^{12} and R^{13} are independently H or C_{1-6} alkyl);

or R¹ and R⁶ form together a linker having 1 to 10 members, wherein the members are selected from C, O, N and S, such that the peptide of Formula I is cyclic;

 R^2 , R^3 , R^4 and R^5 are independently selected from H and C_{1-6} alkyl;

 R^7 , R^8 , R^9 , R^{10} and R^{11} are independently selected from H and methyl;

rings B and C are each independently a 5- to 7-membered saturated, unsaturated or aromatic ring, which may contain one to three heteroatoms selected from O, N and S, and which may be fused to another 5- to 7-membered saturated, unsaturated or aromatic ring;

15 n and p are independently 0, 1 or 2; and wherein

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the stereochemistry at all the α -carbon atoms is L;

and salts and tritiated derivatives thereof, with the exclusion of the compounds:

 $\label{eq:complementary} Ac\text{-Leu-Pro-Phe-Phe-Asp-NH}_2 \text{ and } Ac\text{-Leu-Pro-}(\alpha Me) Phe\text{-Phe-Asp-NH}_2.$

In a second aspect, the invention provides a pharmaceutical composition comprising a compound of Formula I, together with a pharmaceutically acceptable excipient or carrier.

In a third aspect, the invention provides a use of a compound of Formula I for the preparation of a medicament for the treatment or prevention of a disease or condition selected from Alzheimer's disease, Down's syndrome, Dementia pugilistica (including

head trauma), Hereditary Cerebral Haemorrhage with amyloidosis of the Dutch type (HCHWA-D) and vascular dementia with amyloid angiopathy.

In a fourth aspect, the invention provides a use of a compound of Formula I for the treatment or prevention of a disease or condition selected from Alzheimer's disease, Down's syndrome, Dementia pugilistica (including head trauma), Hereditary Cerebral Haemorrhage with amyloidosis of the Dutch type (HCHWA-D) and vascular dementia with amyloid angiopathy.

In a fifth aspect, the invention provides a method of treating Alzheimer's disease, Down's syndrome, Dementia pugilistica (including head trauma), Hereditary Cerebral Haemorrhage with amyloidosis of the Dutch type (HCHWA-D) and vascular dementia with amyloid angiopathy, comprising administering to a patient in need thereof an effective amount of a compound of Formula I.

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In a sixth aspect, the invention provides a use of a compound of Formula I for the preparation of a medicament for the treatment of a disease associated with abnormal protein folding into amyloid and amyloid-like deposits.

20 In a seventh aspect, the invention provides a use of a compound of Formula I for the treatment of a disease associated with abnormal protein folding into amyloid and amyloid-like deposits.

In an eighth aspect, the invention provides a method of treating a disease associated with abnormal protein folding into amyloid and amyloid-like deposits, comprising administering to a patient in need thereof an effective amount of a compound of Formula I.

In a ninth aspect, the invention provides a method for the preparation of a compound of Formula I.

Brief description of the drawings

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Figure 1 shows the *in vitro* activity of compounds of the invention using a fibrillogenesis assay. The compounds were screened for activity using an assay based on quantitating amyloid fibril formation of synthetic A β 1-42. A β (110 μ M) in 0.1 M tris, pH7.4 was incubated alone or in the presence of a ten-fold molar excess of each of the compounds during 5 days at 37°C. Thereafter, the amount of amyloid fibrils was measured using the ThT fluorescent method, as described in the Examples. Values correspond to the percentage of inhibition of amyloid formation and expressed relative to the activity of the comparative compound of Example 15 (100%).

Figure 2 shows the *in vitro* activity of compounds of the invention using a cellular assay. Aβ1-42 (110 μM) in 0.1 M tris, pH 7.4 was incubated alone or in the presence of a ten-fold molar excess of each of the compounds, for 36h at 37°C. The solution was then diluted in cell culture medium and added to semi-confluent PC12 cells. Cells were incubated for 24h and cellular viability was estimated by an MTT assay. Values correspond to the percentage of inhibition of amyloid cytotoxicity and expressed relative to the activity of the comparative compound of Example 16 (100%).

Figure 3 shows a graph representing the calculated % of injected dose of compound of Example 15 taken up per g of brain vs. time. Mice are injected with 0.2 ml of lactate Ringer's solution containing 1% BSA and tritium labelled peptide ("hot") of example 15 (100 000cpm/ml of tritium radioactive peptide).

Figure 4A shows the *in vivo* activity of the compound of Example 1 and the comparative compound of Example 16 in a rat model of cerebral amyloidosis. A representative image shows the difference in amyloid size between the groups of animals treated with vehicle, the comparative compound of Example 16 and the compound of Example 1.

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Figure 4B shows a graph depicting the amyloid area in the different animals as estimated by image analysis after immunohistochemistry.

10 Detailed description of the invention

The compounds of the invention are β -sheet breaking peptides with improved pharmacological profile over known β -sheet breaking peptides.

- β-sheet breaking activity can be detected using, for example, an *in vitro* assay, such as that described by Soto *et al.*⁴³, which measures the ability of test compounds to prevent amyloid fibril formation. Results for compounds of the invention are reported in the Examples.
- Amyloid fibrils are cytotoxic, inducing cell death by apoptosis.⁴⁴ Compounds of the invention can be tested for their ability to prevent cell death induced by amyloid fibrils. Results are reported in the Examples.
- The compounds were also tested using an *in vivo* assay, in which inhibition of amyloid deposition in the brain of animals injected with A\beta 1-42 by compounds of the invention is measured.

A compound having an improved pharmacological profile is considered to be a compound having an increased *in vitro* activity, as measured by either or both of the *in vitro* assays described herein, an increased half-life ($t_{1/2}$) in plasma and/or in brain homogenate, or an increased ability to prevent amyloid deposition *in vivo* in rat brain, as compared with known compounds. "Improved" encompasses those compounds showing an increase in any one of these parameters, or in more than one. Preferably the improvement will be a statistically significant one, preferably with a probability value of < 0.05. Methods of determining the statistical significance of results are well known and documents in the art, and any appropriate method may be used.

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In a preferred group of compounds of Formula I, R^1 is selected from formyl, acetyl, propionoyl, butyroyl, aminoacetyl, methylaminoacetyl, dimethylaminoacetyl, aminoethyl, methylaminoacetyl, dimethylaminoacetyl, preferably R^1 is selected from acetyl, methylaminoacetyl, dimethylaminoacetyl, most preferably acetyl.

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In another preferred group of compounds of Formula I, R⁶ is NHMe or NH₂, particularly preferably NH₂.

In another preferred group of compounds of Formula I, R², R³, R⁴ and R⁵ are selected from H, methyl and ethyl, particularly preferably H and methyl. Particularly preferably R³ is methyl. In a particularly preferred group of compounds, R³ is methyl and R², R⁴ and R⁵ are H.

In another preferred group of compounds of Formula I, R⁹ is methyl and R¹⁰ is methyl or H.

In a particularly preferred group of compounds, R⁹ is methyl, R¹⁰ is methyl or H, and R⁷, R⁸, and R¹¹ are H. In another preferred group of compounds of the invention, R⁷, R⁸, R⁹, R¹⁰ and R¹¹ are H.

In another preferred group of compounds of Formula I, rings B and C are phenyl.

In another preferred group of compounds of Formula I, n and p are both 1.

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The compounds of the invention may be isolated and purified as salts. Such salts fall within the scope of the invention. For the purposes of administration to a patient, it is desirable that the salts be pharmaceutically acceptable.

"Pharmaceutically acceptable salts" refers to salts of the compounds of Formula I that retain the desired biological activity. Examples of such salts include, but are not restricted to, acid addition salts formed with inorganic acids (e.g. hydrochloric acid, hydrobromic acid, sulfuric acid, phosphoric acid, nitric acid, and the like), and salts formed with organic acids such as acetic acid, oxalic acid, tartaric acid, succinic acid, malic acid, fumaric acid, maleic acid, ascorbic acid, benzoic acid, tannic acid, pamoic acid, alginic acid, polyglutamic acid, naphthalene sulfonic acid, naphthalene disulfonic acid, and polygalacturonic acid. Said compounds can also be administered as pharmaceutically acceptable quaternary salts known by a person skilled in the art, which specifically include the quaternary ammonium salts of the formula –NR,R',R" + Z', wherein R, R', R" is independently hydrogen, alkyl, or benzyl, and Z is a counter ion, including chloride, bromide, iodide, alkoxide, toluenesulfonate, methylsulfonate, sulfonate, phosphate, or carboxylate (such as benzoate, succinate, acetate, glycolate, maleate, malate, fumarate, citrate, tartrate, ascorbate, cinnamate, mandelate, and diphenylacetate).

When employed as pharmaceuticals, the compounds of the invention are typically administered in the form of a pharmaceutical composition. Such compositions can be prepared in a manner well known in the pharmaceutical art and comprise at least one active compound. Generally, the compounds of the invention are administered in a pharmaceutically effective amount. The amount of the compound actually administered

will typically be determined by a physician, in the light of the relevant circumstances, including the condition to be treated, the chosen route of administration, the actual compound administered, the age, weight, and response of the individual patient, the severity of the patient's symptoms, and the like.

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The pharmaceutical compositions of the invention can be administered by a variety of routes including oral, rectal, transdermal, intrathecal, subcutaneous, intravenous, intramuscular, and intranasal. Preferably the compounds of the invention are administered by subcutaneous, intramuscular or intravenous injection or infusion.

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In a preferred embodiment of the invention, a compound of the invention is fused to a carrier molecule, a peptide or a protein that promotes the crossing of the blood brain barrier ("BBB"). This serves for proper targeting of the molecule to the site of action in those cases, in which the CNS is involved in the disease. Modalities for drug delivery through the BBB entail disruption of the BBB, either by osmotic means or biochemically by the use of vasoactive substances such as bradykinin. Other strategies to go through the BBB may entail the use of passive diffusion and the use of endogenous transport systems, including carrier-mediated transporters such as glucose and amino acid carriers; receptor-mediated transcytosis for insulin or transferrin; adsorptive-mediated transcytosis. Strategies for drug delivery behind the BBB further include intracerebral implantation.

Depending on the intended route of delivery, the compounds may be formulated as injectable or oral compositions. The compositions for oral administration can take the form of bulk liquid solutions or suspensions, or bulk powders. More commonly, however, the compositions are presented in unit dosage forms to facilitate accurate dosing. The term "unit dosage forms" refers to physically discrete units suitable as unitary dosages for human subjects and other mammals, each unit containing a pre-determined quantity of active material calculated to produce the desired therapeutic effect, in association with a

suitable pharmaceutical excipient. Typical unit dosage forms include pre-filled, pre-measured ampoules or syringes of the liquid compositions or pills, tablets, capsules or the like in the case of solid compositions. In such compositions, the compound of the invention is usually a minor component (from about 0.1 to about 50% by weight or preferably from about 1 to about 40% by weight) with the remainder being various vehicles or carriers and processing aids helpful for forming the desired dosing form.

Liquid forms suitable for oral administration may include a suitable aqueous or non-aqueous vehicle with buffers, suspending and dispensing agents, colorants, flavours and the like. Solid forms may include, for example, any of the following ingredients, or compounds of a similar nature: a binder such as microcrystalline cellulose, gum tragacanth or gelatine; an excipient such as starch or lactose; a disintegrating agent such as alginic acid, Primogel, or corn starch; a lubricant such as magnesium stearate; a glidant such as colloidal silicon dioxide; a sweetening agent such as sucrose or saccharin; or a flavouring agent such as peppermint, methyl salicylate, or orange flavouring.

Injectable compositions are typically based upon injectable sterile saline or phosphatebuffered saline or other injectable carriers known in the art.

The above-described components for orally administered or injectable compositions are merely representative. Further materials as well as processing techniques and the like are known to the skilled practitioner.⁴⁵

The compounds of this invention can also be administered in sustained release forms or from sustained release drug delivery systems. A description of representative sustained release materials is also known to the skilled practitioner.⁴⁶

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The compounds of the invention prevent the aggregation of $A\beta$ associated with the onset and progression of Alzheimer's disease, Down's syndrome, Dementia pugilistica (including head trauma), Hereditary Cerebral Haemorrhage with amyloidosis of the Dutch type

(HCHWA-D) and vascular dementia with amyloid angiopathy. In a preferred method of use of the compounds, administration of the compounds is by injection or infusion, at periodic intervals. The administration of a compound of the invention should preferably begin before any symptoms are detected in the patient, and should continue thereafter. Patients at a high risk for developing Alzheimer's disease, Hereditary Cerebral Haemorrhage with amyloidosis of the Dutch type (HCHWA-D) and vascular dementia with amyloid angiopathy include those with a familial history of these diseases.

Still a further aspect of the present invention is a process for preparing the compounds of Formula I. The compounds of the invention may be prepared from readily available starting materials using the following general methods and procedures. It will be appreciated that where typical or preferred experimental conditions (i.e., reaction temperatures, time, moles of reagents, solvents, etc.) are given, other experimental conditions can also be used unless otherwise stated. Optimum reaction conditions may vary with the particular reactants or solvent used, but such conditions can be determined by one skilled in the art by routine optimisation procedures.

The compounds of the invention may be prepared using methods of peptide synthesis known to the skilled practitioner.⁴⁷ In a preferred embodiment, the compounds of the invention are synthesised using solid-phase methods.

A preferred route to the compounds of the invention is depicted in Scheme 1, and particular examples are given in the Examples that follow.

25 Abbreviations

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The following abbreviations are hereinafter used in the accompanying examples:

min (minute), hr (hour), g (gram), mmol (millimole), m.p. (melting point), eq (equivalents), ml (milliliter), µl (microliters), ACN (acetonitrile), BOP (benzotriazol-1-yl-oxcy-tris-

(dimethylamino)-phosphonium hexafluorophosphate), BOP-Cl (bis(2-oxo-3-oxazoldinyl) Boc (butoxycarbonyl), Cbz (carboxybenzyl), phosphinic chloride), **DCM** (dichloromethane), DIC (diisopropyl carbodiimide), DCC (dicyclohexylcarbodiimide), DIEA (diisopropyl ethylamine), **DMAP** (4-dimethylamino-pyridine), **DMF** (dimethylformamide), EDC (1-(3-dimethylaminopropyl)-3-ethyl-carbodiimide hydrochloride), EtOAc (ethyl acetate), Et₂O (diethylether), Fmoc (9-fluorenylmethoxycarbonyl), (0-(7-azabenzotriazol-1-yl)-1,1,3,3-tetramethyluronium hexafluorophosphate), HOBt (1-hydroxybenzotriazole), rt (room temperature), PyBOP (benzotriazol-1yloxy)tripyrrolidino-phosphonium hexafluorophosphate), PyBrop (bromo-tris-pyrrolidinophosphonium hexafluorophosphate), TBTU (O-benzotriazolyl-N,N,N,N,N-tetramethyl-10 uronium-tetrafluoroborate), TEA (triethylamine), TFA (trifluoro-acetic acid), THF (tetrahydrofuran), cpm (counts per minute), Ci (Curies).

N(Me), in the peptide sequence stands for a methyl group branched on the N atom of the amino acid, (aMe)- in the peptide sequence stands for a methyl group branched on the C alpha of the amino acid.

Synthesis of compounds of the invention:

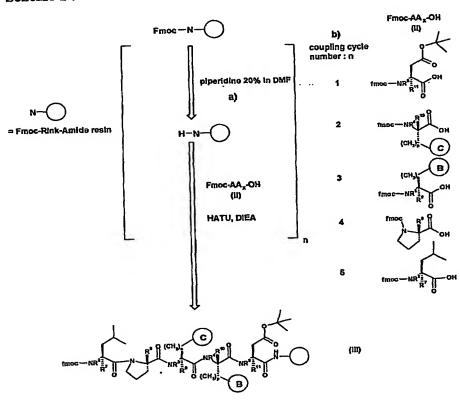
The compounds according to formula I may be prepared from readily available starting materials. Examples of synthetic pathways will be described below.

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General protocol:

A preferred pathway for preparing pentapeptides according to the general Formula I, wherein R¹, R², R³, R⁴, R⁵, R⁶, R⁷, R⁸, R⁹, R¹⁰, R¹¹, B, C, n and p are defined above is described in scheme 1.

Scheme 1:



5 In a preferred embodiement the peptides of formula (I) may be synthesized on a solid support using, for example, preferred Rink-Amide resin.

Typically the resin is washed with DCM, DMF, THF and again DCM three times.

a) Resin deprotection step:

A solution of piperidine in DMF (20-50%) is applied to remove the Fmoc-protecting group from the resin. The resin is gently shacked for 30 minutes to 1hour in such a solution. After draining, the resin is washed with DCM, DMF, THF and again DCM each three times.

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b) Coupling step:

A solution of Fmoc-aspartic acid(β -O-t-butyl) and the resin are shaken with 1.5 equivalents of coupling reagent, such as PyBOP, TBTU, HATU, BOP-Cl, PyBrop, BOP, EDC, DIC, DCC, preferably HATU, and 3 equivalents of a tertiary base, such as NMP, triethylamine, diisopropyl-ethylamine or any tertiary base with similar pKa, preferably diisopropylethylamine. After 2 to 15 hours at between 25 to 40°C, the resin is drained and washed with DCM, DMF, THF and again DCM three times.

For the remaining four amino acids the deprotection and coupling steps (Scheme I, coupling cycles 2 to 5) are applied sequentially in the same manner as described above, using the appropriate Fmoc-protected amino acid of formula (II) to yield resin-bound compound of formula (III).

After the fifth Fmoc-amino acid-coupling step, the group R¹ is introduced as follows.

c) Introduction of R1:

The Fmoc moiety is removed from compound of formula (III) as described above in the deprotection step and the group R¹ is introduced.

For compounds of formula I in which R¹ is an unsubstituted acyl group (scheme 2), a freshly prepared solution of R¹COCl in pyridine (20 equivalents) is shaken with the resinbound compound (compound of formula III) for two to 20 hours at 25 to 40°C (scheme 2).

The resin is extensively washed with DCM, DMF, THF and again DCM three times before being submitted to the cleavage step. The resulting compound wherein the R¹ moiety is present is described by formula (IV).

5 Scheme 2:

d) Cleavage step:

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A freshly prepared solution of 95% TFA in DCM is shacken with the resin for one to four hours at room temperature. This cleaves the compound from the resin as a C-terminal amide and removes the TFA-sensitive protecting groups. The solution is collected and the treatment is repeated two times. The solutions are collected and evaporated to dryness. The crude compound of formula (I) is purified by preparative HPLC using conditions as described below.

For compounds of formula I wherein R1 is methylamino acetoyl or dimethylamino acetoyl, the N-term is blocked by Ac-methyl (di-methyl) amine (R¹), the person skilled in the art may use the same process as described above but step c) is varied as shown on scheme 3.

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Scheme 3:

First, the Fmoc moiety is removed from compound of formula (III) as described above in the deprotection step and the group R^1 is introduced as follows:

Freshly prepared solution of either Boc-sarcosine or N;N-dimethyl-glycine with 1.5 equivalent of coupling reagent, like PyBOP, TBTU, HATU, BOP-Cl, PyBrop, BOP, EDC, DIC, DCC, most preferred HATU and 3 equivalents of a tertiary base, like NMP, triethylamine, diisopropylethylamine or any tertiary base with similar pKa, preferred diisopropylethylamine are shaken together with the resin-bound compound (compound of formula III) for 2 to 15 hours between 25 and 40°C (coupling step 6). After draining the resin is washed with DCM, DMF, THF and again DCM three times. A compound of formula (V) or (VI) is obtained.

The crude compound of formula I wherein R¹ is CH₃NHCH₂C(O)- or (CH₃)₂NCH₂C(O)- (formulae VII and VIII) is obtained after cleavage from the resin as described in d) above and purified by preparative HPLC using conditions as described below.

When R¹ is H₂N(CH2)₂- or (CH3)₂N(CH2)₂-, resin-bound compound of formula III, is treated by reductive amination with Boc-Gly-CHO or Fmoc-Sar-CHO as shown in scheme 4, after removing the Fmoc group as previously described.

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Scheme 4:

In case the case of a Boc-N-Alkyl-leucine is used as terminal amino acid in the final coupling step c) is omitted. The peptide is directly cleaved from the resin as mentioned in step d).

5 Tritiation protocol:

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Tritiated derivatives of compound of formula I can be prepared following the general protocol coupling a 3,4-dehydro-proline residue in step 4. The Dehydro-Pro peptide (2 mg) is mixed with 10% palladium on calcium carbonate (10 mg) and DMF (1 ml) in a tritiation vessel. The mixture is stirred under tritium gas (5 Ci) for 3 hours. The solution is filtered and labile tritium removed by repeated evaporations to dryness with ethanol.

Crude reaction mixture was purified by high performance liquid chromatography in the following system: On a Vydac C18 Protein and Peptide column (250 x 9.6 mm) using a water: acetonitrile:TFA gradient system. The detection is performed by a radioactive detector and a UV detector (220 nm). Product collected, evaporated to dryness, re-dissolved in dispensing eluent.

The other purifications are performed as followed: Preparative HPLC Waters Prep LC 4000 System equipped with columns Prep Nova-Pak®HR C186 µm 60Å, 40x30mm (up to 100mg) or 40x300 mm (up to 1g). All the purifications were performed with a gradient of MeCN/H₂O 0.09% TFA.

The following building blocks are commercially available from Bachem, Switzerland, Fmoc-L-phenylalanine, Fmoc-(β-OtBu)-L-aspartic acid, Fmoc-L-leucine, Fmoc-L-proline, Fmoc-N-Me-L-phenylalanine, Fmoc-N-Me-(OtBu)-L-aspartic acid, Fmoc-α-Me-phenylalanine, Fmoc-N-Me-L-leucine, and Boc-sarcosine. N, N-dimethylglycine is commercially available from Fluka, Switzerland. Boc-Gly-CHO was prepared from Boc-Gly-OH (5.5 mmoles) using a method as described in the literature⁴⁸. Fmoc-Sar-CHO was prepared from

Fmoc-Sar-OH (16 mmoles) using the method as cited above. 3, 4-dehydro-proline is available from Fluka, Switzerland.

Coupling reagents are commercially available from Novabiochem, Switzerland.

Generally, the peptide derivatives according to the general formula (I) can be synthesized using standard peptide synthesis techniques either in solution or on solid phase. In both approaches typical coupling reagents are used, which are known to the person skilled in the art.

It will be also appreciated that where typical or preferred experimental conditions (i.e. reaction temperatures, time, moles of reagents, solvents etc.) are given, other experimental conditions can also be used unless otherwise stated. Optimum reaction conditions may vary with the particular reactants or solvents used, but such conditions will be appreciated by the person skilled in the art.

Examples

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15 The compounds of Examples 1 to 14 are preferred embodiments of the invention:

Compounds of examples 1, 2, 3, 4, 5, 6,7, 8 and 13 may be synthesized according to protocol presented in scheme 1. In case the case of compound of example 13, N-BocMe Leu is the N-terminal amino acid residue and the compound is cleaved from the resin after the final coupling step.

For the synthesis of compound of example 9, the general protocol can be followed using Boc-sarcosine (see scheme 2 in step c). The cleavage step d) is carried out as described in the example above.

For the synthesis of compound of example 10, the general protocol can be followed using N; N-dimethyl-glycine (see scheme 2 in step c).). The cleavage step d) is carried out as described in the example above.

For the synthesis of compound of example 11, the general protocol can be followed and step c) is carried out as described in scheme 3 using Boc-Gly-CHO. Boc-Gly-CHO was dissolved in AcOH/DMF (0.07 ml/7 ml) and was poured into the reaction vessel containing peptide resin (1 mmole). 3 mmol of NaBH₃CN in DMF (3 ml) was added and the reaction mixture was stirred for 5 h. The peptide resin was washed three times with DMF.

For the synthesis of compound of example 12 the general protocol can be followed and step c) is carried out as described in scheme 3 using Fmoc-Sar-CHO. Fmoc-Sar-CHO was dissolved in AcOH/DMF (0.14 ml/14 ml) and was poured into the reaction vessel containing peptide resin (1 mmole). 3 mmoles of NaBH₃CN in DMF (3 ml) was added and the reaction mixture was stirred for 5 h. The peptide resin was washed three times with DMF.

Compound of example 14 is synthesized as follows:

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N-Fmoc-STritylCysteine is coupled to the resin using a coupling reagent as described in scheme 1, step b). The remaining amino acid residues are coupled sequentially according to scheme 1, step b). After the coupling step with Fmoc-Leucine and deprotection with piperidine in DMF, an additional coupling step is carried out with N-Fmoc-S-Trityl Cysteine. The Fmoc group is removed with piperidine in DMF, cleavage and removal of O-Boc and S-trityl groups is accomplished with TFA in DCM. The resulting compound Cys-Leu-Pro-Phe-Asp-Cys is cyclised to form a cyclic disulfide under the following conditions. The peptide is dissolved at 0.1g/l in water. The pH is adjusted to 8.5-9 by addition of Tris HCl buffer at 10 mM. Oxidation is triggered by adding 20% DMSO. The mixture is incubated 24 hours. Oxidation is controlled by mass spectrometry. The crude is purified by HPLC.

Compound of example 15 stands for tritiated compound of example 1 which may be synthesized according to protocol presented in scheme 1 using a 3,4-dehydro-proline amino

acid instead of Proline in step 4. The peptide is then tritiated according to the tritiation protocol above. Compound of example 15 had the specific activity of 42 Ci/mmol.

Example	Structure
1	NH N
2	THE HOLL HAVE
3	JH JOH NH ₂
4	OH OH NH2

Example	Structure
5	JA NATION ON NATIONAL
6	DH OH NH2
7	D N N N N N N N N N N N N N N N N N N N
8	H OH NH2
9	H H H H H NM2

Example	Structure
10	N H NH.
11	H ₂ N — H — NH ₃
12	N N N N N N N N N N N N N N N N N N N
13	NH ₂

Example	Structure
14	

Example N°	Mass Theor.	Mass Exp.
1	692.8	692.3
2	692.8	692.3
3	706.8	706.8
4	706.8	707.4
5	706.8	706.5

		
Example N°	Mass Theor.	Mass Exp.
6	720.9	720.4
7	692.8	693.5
8	706.9	707.4
10	721.08	721.7
11	679.8	679.6
12	707.8	707.5
13	650.8	650.5
··· 14 · ·	···· 842:0 · ··	···· 842.3
15	690.8	691.5

Comparative Example 16

The following compound is disclosed in WO 01/34631 (Axonyx Inc.), and is included as a reference compound.

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Example 16:

10 Example 17: Biological assays

In vitro assays of peptide stability.

The stability of the compounds of the invention can be assayed in comparison with the reference compound (example 16).

Peptides were prepared as a 1µg/µl solution in water. 20 µl of the peptide solution was diluted in 80 µl of fresh human plasma or 10% rat brain homogenate (prepared in PBS). The resulting solution was incubated at 37°C for different times and the reaction was stopped by adding a complete cocktail of protease inhibitors. The bulk of plasma and brain proteins (but none of the peptide) were precipitated in cold methanol (mix/MeOH, 4/5, v/v) during one hour at -20°C. The precipitated proteins were pelleted by centrifugation (10000g, 10 min, 4°C). The supernatant, containing the peptide, was concentrated 5 times under vacuum and separated by reverse-phase HPLC. The area of the peak corresponding

to the intact peptide was measured and compared with an equivalent sample incubated in PBS. The results of are listed in Table 1 as "t₁ human plasma" and "t₂ rat brain homogenate". The values compare favourably with those obtained for the reference compound of Example 16.

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Table 1.	Table 1. In vitro half-lives of various peptides		
Example nº	t _% human plasma	t _{1/2} rat brain homogenate	
1	>24h	>24h	
4	>24h	5h	
5	>24h	>24h	
6	>24h	>24h	
7	>24h	<5 min	
8	>24h	>24h	
9	7h36	41 min	
10	6h	25 min	
16	>24h	15 min	

In vitro assays of activity.

The activity of the compound of the invention in inhibiting the formation of aggregated fibrils can be tested by following the changes in fluorescence signal of a fluorophore that has an affinity for the amyloid fibrils.

Amyloid formation was quantitatively evaluated by the fluorescence emission of thioflavine T (ThT) bound to amyloid fibrils, as reported by Levine⁴⁹ and also Soto *et al.*⁵⁰ Aliquots of A β 1-42 (a fragment of the amyloid beta-peptide known to be amyloidogenic) at

a concentration of 0.5 mg/ml prepared in 0.1M Tris, pH 7.4 were incubated for 5 days at 37°C, gently swirled on a rotary shaker, in the absence or in the presence of different concentrations of the compounds. At the end of the incubation period, 50 mM glycine, pH 9.2 and 2 µM ThT were added in a final volume of 2 ml. Fluorescence was measured at excitation 435 nm and emission 485 nm in a Perkin Elmer, model LS50B fluorescence spectrometer. Using the analytical method⁵¹, the percentage of inhibition of fibril formation caused by compounds of the invention can be calculated. The results are listed in Table 2 and shown graphically in Figure 1. Values correspond to the percentage of inhibition of amyloid formation and are expressed relative to the activity of the compound of Example 16 (100%).

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Table 2. Inhibition of A β 1-42 fibril formation by compounds of the invention		
Example n°	% Inhibition (Compound of Example 16 as 100%)	
1	88	
2	65	
3	40	
4	80	
5	80	
6	85	
7	88	
8	95	
9	90	
10	100	
11	5	
12	45	

Table 2. Inhibition of A β 1-42 fibril formation by compounds of the invention	
Example n°	% Inhibition (Compound of Example 16 as 100%)
13	· 40
14	1

Cellular assay of activity.

5 Amyloid fibrils are cytotoxic, inducing cell death by apoptosis.⁵² The ability of the compounds of the invention in preventing the amyloid formation can be evaluated by measuring the inhibition of the cytotoxicity in a cell assay.

Aliquots of A β 1-42 at a concentration of 0.5 mg/ml prepared in 0.1M Tris, pH 7.4 were incubated alone or in the presence of different concentrations of the compounds for 36h at 37°C, gently swirled on a rotary shaker. At the end of the incubation period, an aliquot of the solution was added to the medium of PC12 cells to reach a final concentration of A β of 55 μ M. The cells were incubated for 24h and thereafter the cellular viability was evaluated using the MTT kit (Roche, Mannheim, Germany) according to the manufacturer's instructions. The results, listed in Table 3, and shown graphically in Figure 2, are reported as the percentage of inhibition of amyloid cytotoxicity, versus A β incubated alone and expressed relative to the activity of the compound of Example 16 (100%).

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Table 3. Inhibition of amyloid cytotoxicity for compounds of the invention	
Example n°	% inhibition of cell death (Compound of Example 16 as 100%)
1	140
6	20
8	5

Blood-brain barrier permeability studies.

The ability of compound of the invention to cross the BBB can be checked by capillary depletion experiments and the kinetics of their penetration into the brain penetration can be measured.

10 a) Capillary depletion and blood washout

Capillary depletion is used to assess the penetration into the brain of compounds of the invention. A "wash-out" step removes blood from the brain so that levels of the compounds of the invention present in the brain parenchyma can be measured.

Capillary depletion studies⁵³ were done in male CD-1 mice (28-36g). Mice are anaesthetized with i.p. urethane (40%) and the left jugular vein is exposed. A tritium labelled peptide of example 15 is injected i.v.. Before sacrificing the animals, blood is collected from the carotid artery (group 1) or from the descending aorta (group 2). After collection of blood, mice of group 1 are sacrificed or blood is removed by injecting 20 ml lactated Ringer's solution (7.19 g/l NaCl, 0.3 g/l KCL, 0.28 CaCl2, 2.1 g/l NaHCO₃, 0.16+

g/l KH₂PO₄, 0.37 g/l MgCl₂ 6H₂O, 0.99 g/l D-glucose, 10 g/l bovine serum albumin, pH 7.4) into the left ventricle of the heart for 30 sec, which removes more than 95% of the vascular contents of the brain (blood brain washout, group 2).

The brain/serum ratio (µl/g) is evaluated by the equation: Brain/serum ratio = (cpm/g brain)/(cpm/µl serum). The cerebral cortex is weighed and homogenized in a physiological buffer (10 mM HEPES, 140 mM NaCl, 4 mM KCl, 2.8 mM CaCl₂, 1 mM MgSO₄, 1 mM Na H₂PO₄, and 10 mM D-glucose, pH 7.4). Dextran solution (1.6 ml of a 26% solution) is then added to the homogenate. After centrifugation (5,400 g, 15 min, 4°C), brain vasculature (pellet) and parenchyma (supernatant) are separated and the radioactivity determined in each fraction.

After 10 min post i.v injection of radioactive compound 1, more than 70% of the radioactivity is recovered in the parenchyma. The percentage of radioactivity associated with the capillary fraction with or without blood wash out is 24 % and 29%, respectively. These data show that the majority of compound 1 crosses the BBB. In addition, it proves that the peptide of example 1 is weakly binding to the luminal surface of the capillaries and is not sequestrated into the endothelial cells.

b) Blood brain barrier permeability study:

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The kinetics of penetration of compound of the invention into the brain can be evaluated through blood brain barrier permeability experiments. The percentage of injected peptide found in the brain can then be calculated.

Mice are anaesthetized with i.p. urethane (40%) and the left jugular vein is exposed. 0.2 ml of lactate Ringer's solution (7.19 g/l NaCl, 0.3 g/l KCL, 0.28 CaCl2, 2.1 g/l NaHCO₃, 0.16+ g/l KH₂PO₄, 0.37 g/l MgCl₂6H₂O, 0.99 g/l D-glucose, 10 g/l bovine serum albumin, pH 7.4) containing 1% BSA and tritium labelled peptide ("hot") of example 15 (100 000cpm/ml of tritium radioactive peptide) is injected. Arterial blood is collected from the

right carotid artery at different time points following the labelled peptide injection. Serum is obtained by centrifugation (4800 g, 10 min, 4°C). Following arterial blood collection, the mice are decapitated and the whole brains, except the pineals and pituitaries, are harvested and weighed. The amounts of radioactivity in brain and serum are determined after an overnight solubilization step in TS-2 solution (RPI, Mount Prospect, IL) at 40°C. The brain serum ratio of total radioactivity was determined over time from 1 min up to 120 min after injection. The brain to serum ratio (μ l/g) is estimated by the equation: Brain/serum ratio= (cpm/g brain)/(cpm/ μ l serum).

The representation of the brain to serum ratio versus time allows the derivation of the influx rate, Ki (slope) and the volume of distribution (Y intercept), Vi. The influx rate (Ki, microl (serum)/g (tissue weight)-min) represents the rate at which compounds move from the circulation to the brain. The volume of distribution (Vi, microl (serum)/g (tissue weight)) is the apparent volume of material which is distributed to the brain parenchyma at time 0 min.

15 Results are presented in Table 4.

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Table 4. Brain up	take study of compound of	Example 15
Injection (i.v.) composition	Ki (microl/g-min)	Vi (microl/g)
Hot*	3.54 +/- 0.29	50.6 +/- 12.4

*Hot: Radioactively labeled compound

Ki: influx rate

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Vi: volume of distribution

The influx rate (Ki), with radioactive peptide, alone was 3.54 +/- 0.29 microl/g-min with a Y intercept of about 50 microl/g (Table 1). This indicates that compound 1 penetrates the brain.

5 The percentage of injected dose of compound of example 15, taken up per g of brain, was calculated (see figure 3). The maximum value was 0.464% and half of this value was reached 1.4 min after injection. This data suggests a rapid brain uptake of compound of example 15.

HPLC analysis of brain samples showed that 12% of intact peptide was recovered 20 min post i.v. injection.

More than 0.06% of the injected compound was recovered intact in the brain within the 20 min post injection.

15 In vivo studies using an animal model of cerebral A β deposition.

The inhibitory activity the compound of the invention in the formation of amyloid deposits by can be visualized *in vivo* by staining cerebral tissue sections with anti-A β 1-42 antibodies in the presence and absence of a peptide of the invention.

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Male Fischer-344 rats weighed 250-300g and were 3-4 months of age at the time of arrival. The animals were housed 2 per cage, maintained on a 12 h light-dark cycle with access to food and water *ad libitum* and were habituated to their new environment for 2-3 weeks prior to surgery. Surgery was performed under sodium pentobarbital (50 mg/kg, i.p.) anaesthesia.

Atropine sulphate (0.4 mg/kg) and ampicillin sodium salt (50 mg/kg) were injected subcutaneously once the animals were anaesthetized. $A\beta 1-42$ was dissolved in dimethylsulfoxide (DMSO) and then diluted with water to a concentration of 16.7%

DMSO. The animal received a bilateral injection of 5.0 nmol A β 1-42 into each amygdale by using a Kopf stereotaxic instrument with the incisor bar set at 3.3 mm below the interaural line. Injection coordinates measured from the bregma and the surface of the skull (AP -3.0, ML \pm 4.6, DV -8.8) were empirically determined based on the atlas of Paxinos and Watson. A volume of 3.0 μ 1 of the solution of A β 1-42 at 5.0 nmol was administered over 6 min (flow rate 0.5 μ 1/min) using a CMA/100 micro syringe pump. The cannula was left *in situ* for 2 min following injection, then it was withdrawn 0.2 mm and left for 3 min, and following these 5 min the cannula was slowly withdrawn. Following surgery the animals were placed on a heating pad until they regained their righting reflex. The animals were treated with compound of example 1 and of example 16. The peptides, solubilized in a 0.9% NaCl at the concentration of 4.4 mM were injected s.c (0.5 ml per injection), 4 times a week during 7 and a half weeks.

After treatment with the compounds, animals were sacrificed by an overdose of sodium pentobarbital (150 mg/kg, i.p.), perfused transacrtically. For histology, serial coronal sections (40 μ m) of the brain were cut, placed in ethylene glycol cryoprotectant and stored at -20°C until stained. Tissue sections were stained with anti-A β 1-42 antibodies, as described. An image analysis system was used to determine the size of the amyloid deposits. These data were analysed by a two-way ANOVA followed by a Newman-Keuls' multiple range test for *post hoc* comparisons.

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The photographic results for the compound of Example 1 in comparison with vehicle and the compound of comparative Example 16 are shown in Figure 4A. Graphical results depicting the amyloid area are shown in Figure 4B. It is clear that the compound of Example 1 substantially reduces the amyloid area.

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We claim:

1. A compound of the general formula I:

$$R^{1}$$

$$NR^{2}$$

$$R^{7}$$

$$R^{8}$$

$$NR^{3}$$

$$R^{8}$$

$$R^{10}$$

$$NR^{5}$$

$$R^{11}$$

$$R^{11}$$

$$R^{10}$$

$$R^{10$$

5 wherein:

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R¹ is selected from C₂₋₆-acyl (optionally substituted by amino, C₁₋₆alkyl amino or di-C₁₋₆alkylamino), and C₁₋₆alkyl (optionally substituted by amino, C₁₋₆alkyl amino or di-C₁₋₆alkylamino);

 R^6 is $NR^{12}R^{13}$ (wherein R^{12} and R^{13} are independently H or C_{1-6} alkyl);

or R¹ and R⁶ form together a linker having 1 to 10 members, wherein the members are selected from C, O, N and S, such that the peptide of Formula I is cyclic;

R², R³, R⁴ and R⁵ are independently selected from H and C₁₋₆alkyl;

R⁷, R⁸, R⁹, R¹⁰ and R¹¹ are independently selected from H and methyl;

rings B and C are each independently a 5- to 7-membered saturated, unsaturated or aromatic ring, which may contain one to three heteroatoms selected from O, N and S,

and which may be fused to another 5- to 7-membered saturated, unsaturated or aromatic ring;

n and p are independently 0, 1 or 2; and wherein

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the stereochemistry at all the α -carbon atoms is L;

and salts and tritiated derivatives thereof, with the exclusion of the compounds Ac-Leu-Pro-Phe-Asp-NH₂ and Ac-Leu-Pro-(\alpha Me)Phe-Phe-Asp-NH₂.

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- 2. A compound according to claim 1, wherein R¹ is selected from C₂₋₆-acyl (optionally substituted by amino, C₁₋₆alkyl amino or di-C₁₋₆alkylamino), and C₁₋₆alkyl (optionally substituted by amino, C₁₋₆alkyl amino or di-C₁₋₆alkylamino).
- 15 3. A compound according to claim 1, wherein R¹ is selected from formyl, acetyl, propionoyl, butyroyl, aminoacetyl, methylaminoacetyl, dimethylaminoacetyl, aminoethyl, methylaminoethyl, dimethylaminoethyl and methyl.
- 4. A compound according to claim 1, wherein R¹ is selected from acetyl, methylaminoacetyl and dimethylaminoacetyl.
 - 5. A compound according to claim 1, wherein R¹ is acetyl.
- 6. A compound according to claims 1 to 5, wherein R⁶ is NR¹²R¹³ (wherein R¹² and R¹³
 25 are independently H or C₁₋₆alkyl).
 - 7. A compound according to any one of the preceding claims, wherein R⁶ is NHMe or NH₂.

- 8. A compound according to any one of the preceding claims, wherein R⁶ is NH₂.
- 9. A compound according to any one of the preceding claims, wherein R², R³, R⁴ and R⁵
 5 are selected from H, methyl and ethyl.
 - 10. A compound according to any one of the preceding claims, wherein R², R³, R⁴ and R⁵ are selected from H and methyl.
- 10 11. A compound according to any one of the preceding claims, wherein R³ is methyl.
 - 12. A compound of claim 11 wherein R2, R4 and R5 are H.

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- 13. A compound of claim 11 wherein R⁴ and R⁵ are methyl and R² is H.
- 14. A compound of claim 11 wherein R^5 is methyl, R^2 and R^4 are H.
- 15. A compound of claim 11 wherein R⁴ is methyl and R² and R⁵ is H.
- 20 16.A compound according to any one of the preceding claims, wherein R¹⁰ is methyl and R⁷, R⁸ and R¹¹ are H.
 - 17.A compound according to any one of the preceding claims, wherein R^{10} and R^9 are methyl and R^7 , R^8 and R^{11} are H.
 - 18. A compound of any claims 1 to 10 wherein R², R³, R⁴, R⁵, R⁷, R⁸, R⁹, R¹⁰ and R¹¹ are H.

- 19. A compound according to any one of the preceding claims, wherein rings B and C are phenyl.
- 20. A compound according to any one of the preceding claims, wherein n and p are both 1.

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- 21. A compound according to any one of the preceding claims selected from the following group:
 - Ac-Leu-Pro-N(Me)-Phe-Phe-Asp-NH₂

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Ac-Leu-Pro-Phe-N(Me)-Phe-Asp-NH₂

Ac-Leu-Pro-Phe-N(Me)-Phe-N(αMe)-Asp-NH₂

15 Ac-Leu-Pro-N(Me)-Phe-Phe-N(αMe)-Asp-NH₂

Ac-Leu-Pro-N(Me)-Phe-N(Me)-Phe-Asp-NH₂

Ac-Leu-Pro-N(Me)-Phe-N(Me)-Phe-N(\alpha Me)-Asp-NH2

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Ac-Leu-Pro-Phe-(\alpha Me)-Phe-Asp-NH2

Ac-Leu-Pro-(\alpha Me)-Phe-(\alpha Me)-Phe-Asp-NH2

25 CH₃NHCH₂C(O)-Leu-Pro-Phe-Phe-Asp-NH₂

(CH₃)₂NHCH₂C(O)-Leu-Pro-Phe-Phe-Asp-NH₂

NH₂(CH₂)₂-Leu-Pro-Phe-Phe-Asp-NH₂

(CH₃)₂N(CH₂)₂-Leu-Pro-Phe-Phe-Asp-NH₂

5 CH₃-Leu-Pro-Phe-Phe-Asp-NH₂

Ac-Leu-[3H]Pro-N(Me)-Phe-Phe-Asp-NH2

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- 22. A compound according to any one of the preceding claims for use as a medicament.
- 23. A pharmaceutical composition comprising a compound according to any one of claims 1 to 21, and a pharmaceutically acceptable excipient, diluent or carrier.

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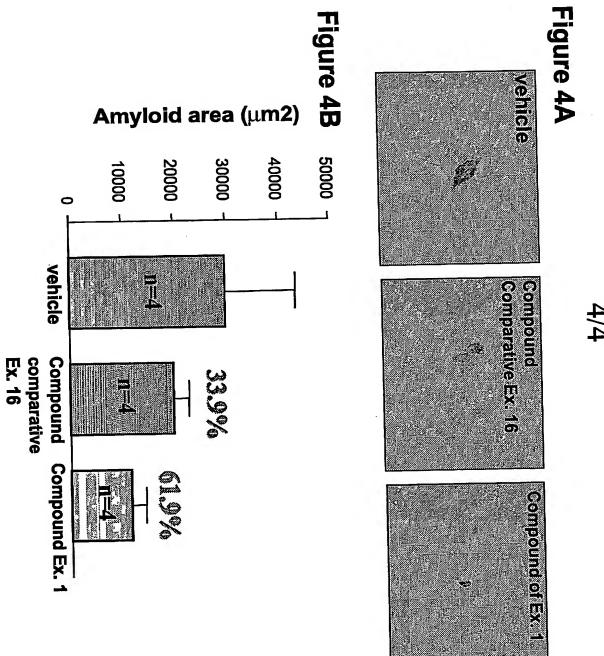
- 24. A use of a compound according to any one of claims 1 to 21 for the manufacture of a medicament for the treatment or prevention of a disease or condition selected from Alzheimer's disease, Down's syndrome, Dementia pugilistica (including head trauma), Hereditary Cerebral Haemorrhage with amyloidosis of the Dutch type (HCHWA-D) and vascular dementia with amyloid angiopathy.
- 25. The use according to claim 24, wherein the disease is Alzheimer's disease.
- 26. A use of a compound according to any one of claims 1 to 21, for the preparation of a medicament for the treatment of a disease associated with abnormal protein folding into amyloid and amyloid-like deposits.

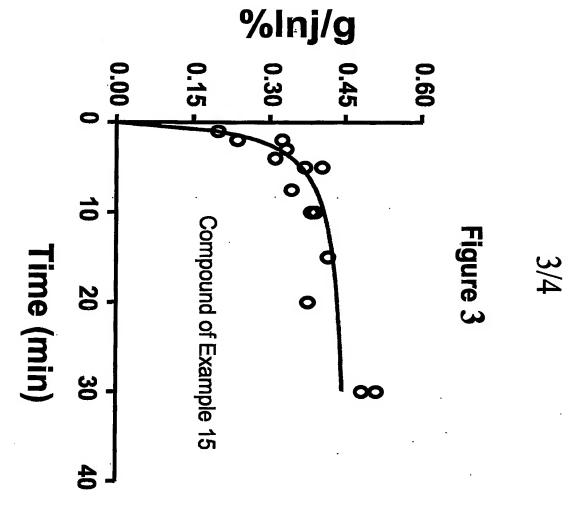
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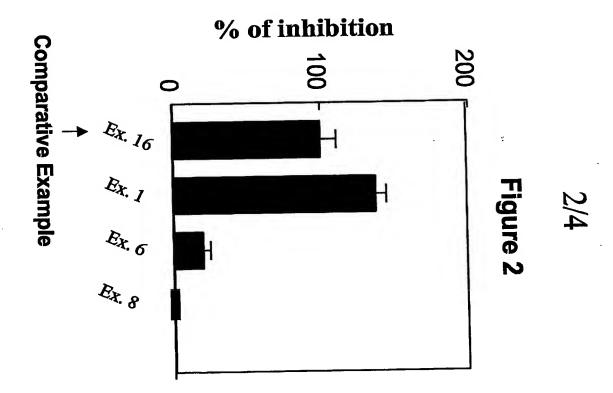
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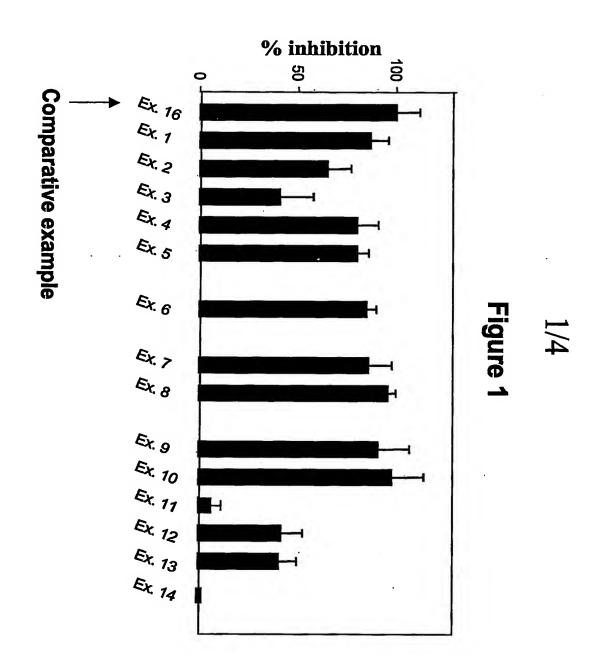
Provided are peptides having β -sheet breaking ability, useful in the treatment and prevention of diseases such as Alzheimer's disease, Down's syndrome, Dementia pugilistica (including head trauma), Hereditary Cerebral Haemorrhage with amyloidosis of the Dutch type (HCHWA-D) and vascular dementia with amyloid angiopathy.

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